



# Evolution of male pregnancy associated with remodeling of canonical vertebrate immunity in seahorses and pipefishes

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**A fundamental problem for the evolution of pregnancy, the most specialized form of parental investment among vertebrates, is the rejection of the nonself-embryo. Mammals achieve immunological tolerance by down-regulating both major histocompatibility complex pathways (MHC I and II). Although pregnancy has evolved multiple times independently among vertebrates, knowledge of associated immune system adjustments is restricted to mammals. All of them (except monotremata) display full internal pregnancy, making evolutionary reconstructions within the class mammalia meaningless. Here, we study the seahorse and pipefish family (syngnathids) that have evolved male pregnancy across a gradient from external oviparity to internal gestation. We assess how immunological tolerance is achieved by reconstruction of the immune gene repertoire in a comprehensive sample of 12 seahorse and pipefish genomes along the “male pregnancy” gradient together with expression patterns of key immune and pregnancy genes in reproductive tissues. We found that the evolution of pregnancy coincided with a modification of the adaptive immune system. Divergent genomic rearrangements of the MHC II pathway among fully pregnant species were identified in both genera of the syngnathids: The pipefishes (*Syngnathus*) displayed loss of several genes of the MHC II pathway while seahorses (*Hippocampus*) featured a highly divergent invariant chain (*CD74*). Our findings suggest that a trade-off between immunological tolerance and embryo rejection accompanied the evolution of unique male pregnancy. That pipefishes survive in an ocean of microbes without one arm of the adaptive immune defense suggests a high degree of immunological flexibility among vertebrates, which may advance our understanding of immune-deficiency diseases.**

immunological tolerance | major histocompatibility complex | male pregnancy | seahorse | comparative genomics

**P**regnancy is the most dramatic form of parental investment, protecting embryos from extreme temperatures, anoxia, osmotic stress, and predation at the cost of fewer young, less effective dispersal, and high-energy demand (1, 2). Although this trait requires multiple anatomical and physiological changes (3) involving development, morphology, osmoregulation, endocrinology, and immunology (4, 5), it has evolved independently in more than 150 vertebrate lineages. A fundamental problem for pregnancy to evolve is the rejection of the embryo that is recognized as foreign tissue by the vertebrate’s adaptive immune system, as it displays alleles also from the other parent. Modulation of the immune system to tolerate foreign protein signatures of the embryonic tissue, in turn, is conflicting with the maintenance of immunological vigilance toward pathogens (6).

Being mammals themselves, researchers have almost exclusively focused on mammalian pregnancy to assess the key adaptations for pregnancy evolution. In vertebrates, the unique diversity of the classic major histocompatibility complex (MHC)

class I and II genes (7–9) plays a key role for self/nonself-recognition. While in mammals an initial inflammation seems crucial for embryo implantation (10), during pregnancy mammals prevent an immunological rejection of the embryo with tissue layers of specialized fetal cells, the trophoblasts (11–13). Trophoblasts do not express MHC II (14–16) and thus prevent antigen presentation to maternal T-helper (Th) cells (17), which otherwise would trigger an immune response against nonself. Additionally, expression of classic *MHC I* genes (HLA-A, -B, and -D) is down-regulated (18). These immunological adaptations are mediated by a cross-talk between the placental trophoblasts and uterine immune cells, in particular natural killer cells and regulatory T cells (Tregs) (19, 20). Tregs maintain self-tolerance by suppressing inflammatory Th1 immune responses

## Significance

Among vertebrates, pregnancy has evolved more than 150 times independently. A fundamental problem for pregnancy to evolve is inadvertent rejection of the embryo when being recognized as foreign tissue by the vertebrate’s adaptive immune system. We show that the unique evolution of male pregnancy in pipefishes and seahorses coincided with a genomic modification of one arm of the adaptive immune system. Our findings indicate a trade-off between immunological tolerance and embryo rejection to accompanying the emergence of male pregnancy. That syngnathids survive in an ocean of microbes despite their drastically modified immune defense suggests an unexpected immunological flexibility. Our results may improve the understanding of immune-deficiency diseases and call for a reassessment of vertebrate immunity.

Author contributions: O.R. and T.B.H.R. initiated and planned the study; S.J. coordinated the sequencing and the comparative genomics; O.K.T. assembled and annotated the genomes and analyzed genome size and repetitive elements; M.H.S. did the gene mining and the alignments, with support from O.R., H.T.B., S.N.K.H., M.S.O.B., and T.B.H.R.; M.M. did the phylogenetic analyses; T.B. did the positive selection analyses; O.R., T.B., and D.H. did the transcriptomics; R.H. provided samples for the genomes; O.R., M.H.S., T.B., T.B.H.R., and S.J. interpreted the data; O.R. wrote the manuscript and made the revisions, with support from all authors.

The authors declare no competing interest.

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Data deposition: The sequences reported in this paper have been deposited in the European Nucleotide Archive, <https://www.ebi.ac.uk/ena> (accession no. PRJEB32126). Gene alignments for positive selection analyses can be found at Figshare: doi:10.6084/m9.figshare.11499360.

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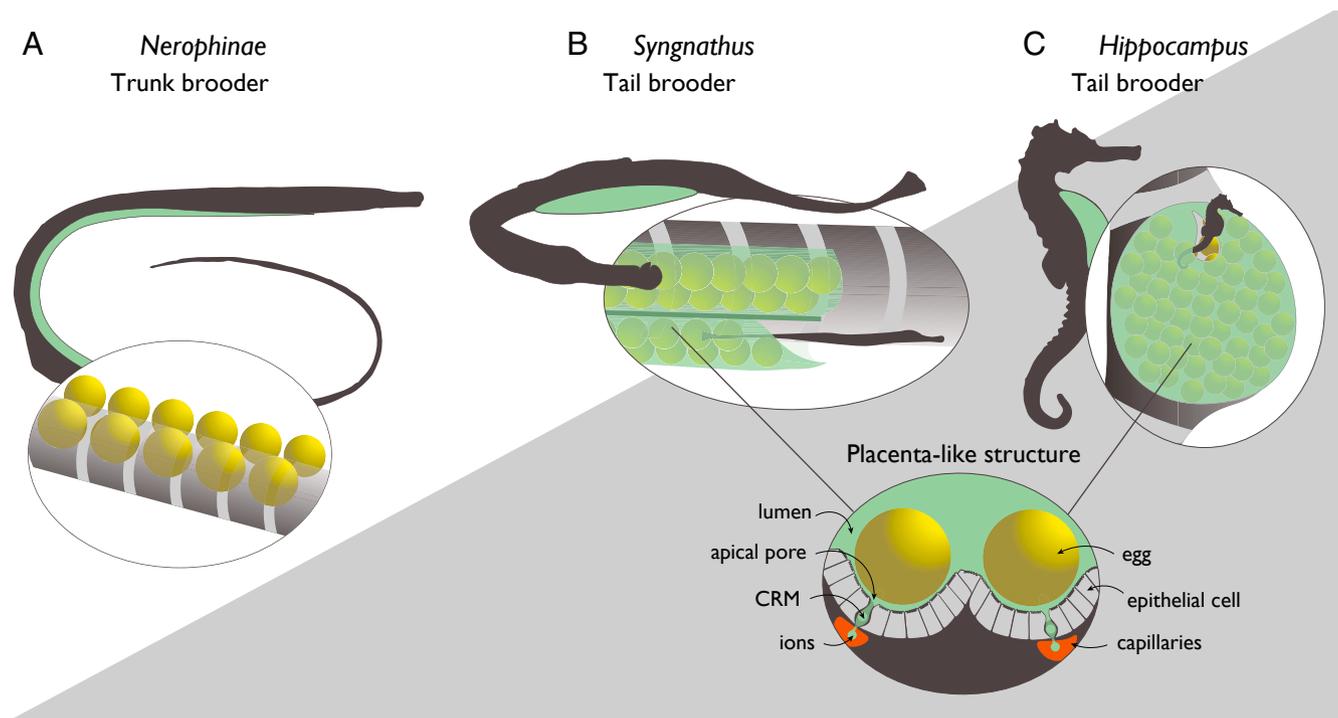
(21), as implied by the fact that Treg deficiencies evoke miscarriage (6).

In order to gain insight into the successive evolution of pregnancy and the corresponding molecular cooption of genes and pathways, comparative studies along well-resolved phylogenetic clades are essential. The lack of transitional stages renders mammals unsuitable to reconstruct critical steps for the evolution of pregnancy. Fishes show the greatest diversity in pregnancy evolution, which makes them pivotal for the assessment of the evolution of vertebrate pregnancy (5). Pregnancy has evolved in six teleost orders (Lophiiformes, Beloniformes, Cyprinodontiformes, Scopaeniformes, Perciformes, and Syngnathiformes) (22). Among those, we selected the Syngnathiformes as an ideal taxonomic group to reconstruct possible genomic modifications underlying pregnancy evolution. Species in this family display a gradient of male pregnancy, ranging from external attachment of the eggs to the belly (in the subfamily *Nerophinae*) to additional external protection via skin flaps (in *Doryrhamphus*, *Oostethus*, and *Solegnathiinae*), with evolution of internal gestation via brood pouches in *Syngnathus* (inverted brood pouch) and *Hippocampus* (sealed brood pouch) as the most advanced states of full internal pregnancy (23, 24) (Fig. 1). In the latter two genera, the fertilized eggs (and later the hatched embryos) become engulfed and effectively integrated by parental tissue, and are supplied with nutrients, oxygen, and parental immunity via a placenta-like organ (1, 25, 26). This gradient in parental investment provides a unique opportunity to analyze the concomitant changes in the vertebrate immune system, and to test whether adaptations similar to mammalian pregnancy assure immunological tolerance. We hypothesized that immunological tolerance toward embryonic tissue is correlated to a genomic modification of the adaptive immune system. We focused particularly on the MHC I and MHC II pathways due to their key roles in immunological tolerance, a hypothesis that was also based on preliminary transcriptomic data in one genus (27).

Here we present comprehensive genome data on 12 representative species of the Syngnathiformes covering a broad range of parental reproductive investment. By assessing their immune gene repertoire, we reconstructed the evolutionary acquisition of immunological tolerance within this unique lineage. As a non-mutually exclusive explanation, we also assessed whether immune gene regulation contributes to immunological tolerance in syngnathids, similar to mammals, and measured differential gene expression during male pregnancy in *Syngnathus typhle* using comparative transcriptomics to assess whether immunological tolerance can also be achieved by gene regulation in syngnathids, similar to mammals.

### Genome Size Evolution in Syngnathiformes

We selected one species, *S. typhle*, to obtain a high coverage and contiguous genome assembled to a high-quality draft stage sufficient to achieve gene repertoire completeness using a combination of paired-end and mate-pair libraries. The genomes of 11 additional species were assembled to draft stage (*SI Appendix, Table S2*). Based on the whole-genome datasets, including the already published genomes of *Syngnathus scovelli* and *Hippocampus comes* (29, 30), Bayesian phylogenetic analyses (*SI Appendix, Table S3*) places the origin of the Syngnathiformes clade at 80 Mya (*SI Appendix, Fig. S1*). Surprisingly, the Syngnathiformes lineage contains species with very divergent genome sizes, spanning from 347 Mbp (*Syngnathus rostellatus*) to 1.8 Gbp (*Entelurus aequoreus*) (Table 1). Syngnathiformes species lacking male pregnancy—namely *Fistularia tabacaria*, *Mullus surmuletus*, *Dactylopterus volitans*, *Aeoliscus strigatus*, and *Macroramphoriscus scolopax*—displayed larger genomes than both genera with full male pregnancy (i.e., all *Hippocampus* and *Syngnathus* species). In contrast, the *Nerophinae* pipefishes with external male pregnancy, specifically *Nerophis ophidion* and *E. aequoreus*, have significantly larger genomes (Table 1). Concordantly, during 50 million y of evolution, transposable elements have expanded in



**Fig. 1.** Morphology of brood pouches of subfamily *Nerophinae* (A), and of the genera *Syngnathus* (B), and *Hippocampus* (C) and display of the placenta-like structures in syngnathids (only *Syngnathus* and *Hippocampus*). The placenta-like structure with lumen, apical pore, CRM (cells rich in mitochondria), ions, epithelial cells, capillaries, and the egg are drawn after figure 1 of ref. 28.

**Table 1. Summary of species, estimated genome sizes, and assembly statistics**

Species	Name	Estimated genome size (Mbp)	Assembly size (Mbp)	N50 scaffold (Mbp)	N50 contig	BUSCO complete (%)
<i>Aeoliscus strigatus</i>	Jointed razorfish	403	381	115.8	15.9	89.6
<i>Dactylopterus volitans</i>	Flying Gurnard	499	577	17.1	8.3	74.1
<i>Doryramphus dactylophorus</i>	Banded pipefish	651	619	75.2	27.6	87.1
<i>Entelurus aequoreus</i>	Snake pipefish	1834	557	3.9	3.4	21.6
<i>Fistularia tabacaria</i>	Bluespotted cornetfish	762	593	107.2	17.7	90.8
<i>Hippocampus comes</i> *	Tiger tail seahorse	NA	494	2034.5	39.6	89.4
<i>Hippocampus kuda</i>	Yellow seahorse	478	445	31.2	10.4	83.9
<i>Hippocampus whitei</i>	White's seahorse	461	433	40.8	10.3	86.0
<i>Macroramphorus scolopax</i>	Longspine snipefish	507	418	41.8	13.4	86.1
<i>Mullus surmuletus</i>	Surmulet	569	469	17.2	7.2	73.8
<i>Nerophis ophidion</i>	Straightnose pipefish	1581	976	6.8	5.2	33.6
<i>Syngnathus rostellatus</i>	Nilsson's pipefish	347	283	87.6	14.9	89.0
<i>Syngnathus scovelli</i> *	Gulf pipefish	NA	307	12400.1	27.8	85.8
<i>Syngnathus typhle</i>	Broadnosed pipefish	NA	315	3047.0	25.8	88.8

NA, not applicable.

\*Already published genomes.

*Nerophinae*, most likely explaining the large genome sizes within this subfamily (*SI Appendix*, Fig. S2 and Table S4).

### Modification of MHC II Pathways in *Syngnathus* and *Hippocampus*

In order to correlate the modification of adaptive immunity with the degree of male pregnancy, a set of key genes involved in adaptive immunity was analyzed from the assembled genomes presented here along with two previously published syngnathid genomes (29, 30) (*SI Appendix*, Table S2). MHC I and MHC II are essential for the recognition process of nonself-peptides by presenting them to CD8<sup>+</sup> and CD4<sup>+</sup> T cells, respectively. In line with our hypothesis, all fully pregnant species (i.e., genera *Syngnathus* and *Hippocampus*) underwent considerable modifications of their adaptive immune system characterized by losses or changes of key genes of the MHC II pathway (Fig. 2; details on ortholog search and analyses can be found in the *SI Appendix*, sections 5.1. and 5.5.11).

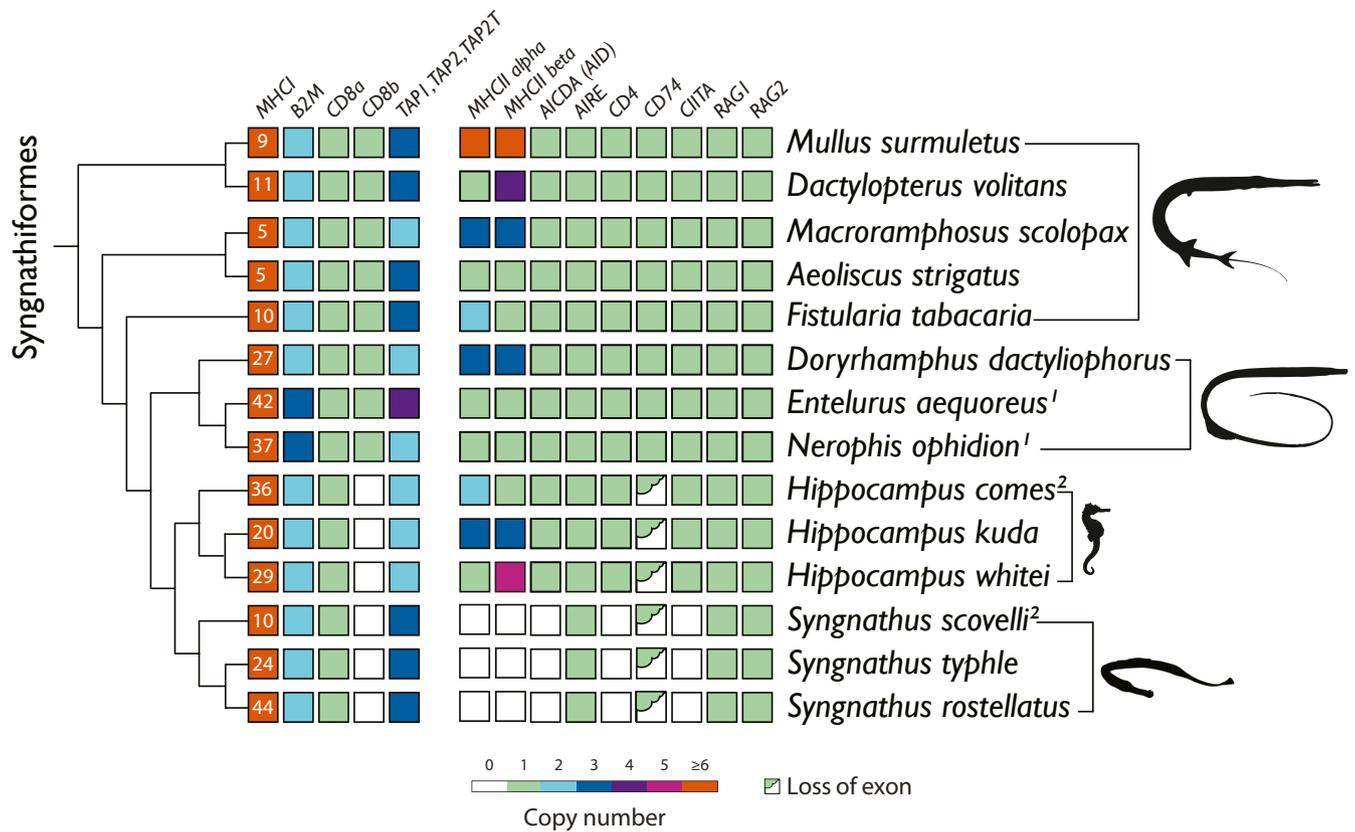
The invariant chain of MHC II (*CD74*), preventing premature peptide binding of MHC II, displayed a divergent exon 3 in *Syngnathus* and *Hippocampus* compared to both mammals and other teleosts (Fig. 3). Additionally, *Hippocampus* had a sequence substitution of exon 6b, while *Syngnathus* displayed a divergent exon compared to other fish and human. Both exons 3 and 6b are located in the protein region protruding into the endosomal lumen. Several lines of evidence suggest that these losses are impairing functions of *CD74*. In human, exon 3 of *CD74* covers the region associating with MHC II (CLIP) [amino acids 108 to 124 in *SI Appendix*, Dataset 1: 10CD74\_clean (31)]. Exon 6b is annotated as Thyroglobulin type I repeats, which are proposed cysteine protease inhibitors (this exon consists of six conserved cysteine residues) and are implicated in delaying the degradation of internalized antigens subsequently preserving epitopes for antigen presentation (32, 33).

As the most drastic change in gene repertoire, all *Syngnathus* species have lost the genes encoding the classic MHC II  $\alpha$ - and  $\beta$ -chains, implying that the presentation of antigens to the T cell receptor on CD4<sup>+</sup> T lymphocytes is disabled (Fig. 2). This is supported by a loss of *CD4*, mediating successful receptor binding and activation of CD4<sup>+</sup> T lymphocytes—*AICDA*, responsible for the unique receptor diversity of the antibodies and *CIITA*, the MHC II transactivator—which control the expression of MHC II genes in antigen-presenting cells. The only canonical gene of the

MHC II pathway remaining in the *Syngnathus* genomes was the auto-immune regulator (35), driving negative selection on self-recognizing T cells (36). While leading and trailing exons of *AIRE* were well conserved among all investigated Syngnathiformes species compared to reference sequences from other fish families, several other exons diverged markedly or homologous sequences were not found [Fig. 3 and *SI Appendix*, Dataset 1: 13AIRE\_exon\_overview (31)]. In the genera *Syngnathus* and *Hippocampus*, exons 3, 4, 5, 6, and 12 of *AIRE* were lost or substituted with very divergent sequences that could not be aligned. Putative loss of MHC II-related function of the *AIRE* transcription factor is further emphasized by the lack of expression in various *S. typhle* tissues, which could result in insufficient negative selection of T cells in the thymus (36). Overall, our findings suggest that the MHC class II pathway was lost in *Syngnathus*.

The situation in *Hippocampus* was more complex. Similar modifications as in *Syngnathus* for the *CD74* gene were observed in terms of a divergent exon 3, and in a substitution of exon 6b. Importantly, no loss of the MHC II genes as in all three *Syngnathus* species was observed. However, in *Hippocampus*, the MHC II gene sequences, in particular of the  $\beta$ -copy, were highly distinct from other functional MHC II genes found in species with functional MHC class II such as zebrafish, seabass, salmon, and guppy [*SI Appendix*, Dataset 1: 29\_MHCII\_beta\_complete (31)]. In parallel, small sections of some *CIITA* exons displayed substitutions compared to both other teleosts and mammals. This is in line with findings for *AIRE* in both *Hippocampus* and *Syngnathus*, where several exons were either lost or diverged markedly compared to other teleosts, indicating most likely an alternative function not related to MHC II (Fig. 3). Moreover, the tertiary structure of MHC II  $\beta$  genes of *Hippocampus* was predicted to lack two critical cysteine bridges that are essential to form the peptide-binding pocket of the MHC II molecule [*SI Appendix*, Dataset 1: 29MHCII\_beta\_complete (31)]. In line with these findings, in *Hippocampus* we identified positive selection in sequences of genes that were lost in *Syngnathus* (*AIRE*, *CD4*, and *CIITA*), which may suggest neo- or subfunctionalization (*SI Appendix*, Table S11).

A closer examination of the invariant chain encoding gene *CD74* also suggests that the evolution of adaptive immunity has taken distinct routes in the two sister genera *Syngnathus* and *Hippocampus*. A shared relaxed selection on *CD74* in the common ancestor of syngnathids resulted in a loss-of-function by either sequence substitution in the *Hippocampus* or divergence



**Fig. 2.** Remodeling and loss of key genes of the MHC class I and II pathways among 14 species of Syngnathiformes. Species are arranged according to their phylogeny (SI Appendix, Fig. S1). The copy numbers of the most important genes of the MHC I pathway (*MHC I*, *B2M*, *CD8a*, *CD8b* and *TAP1*, *TAP2*, *TAP2T*) and MHC II pathway (*MHC II $\alpha$* , *MHC II $\beta$* , *AICDA* [AID], *AIRE*, *CD4*, *CD74*, *CIITA*, *RAG1*, *RAG2*) are displayed. Superscript “1” specifies large fragmented genomes; superscript “2” specifies previously published genomes. White boxes indicate loss of genes and half green boxes indicate loss of exons.

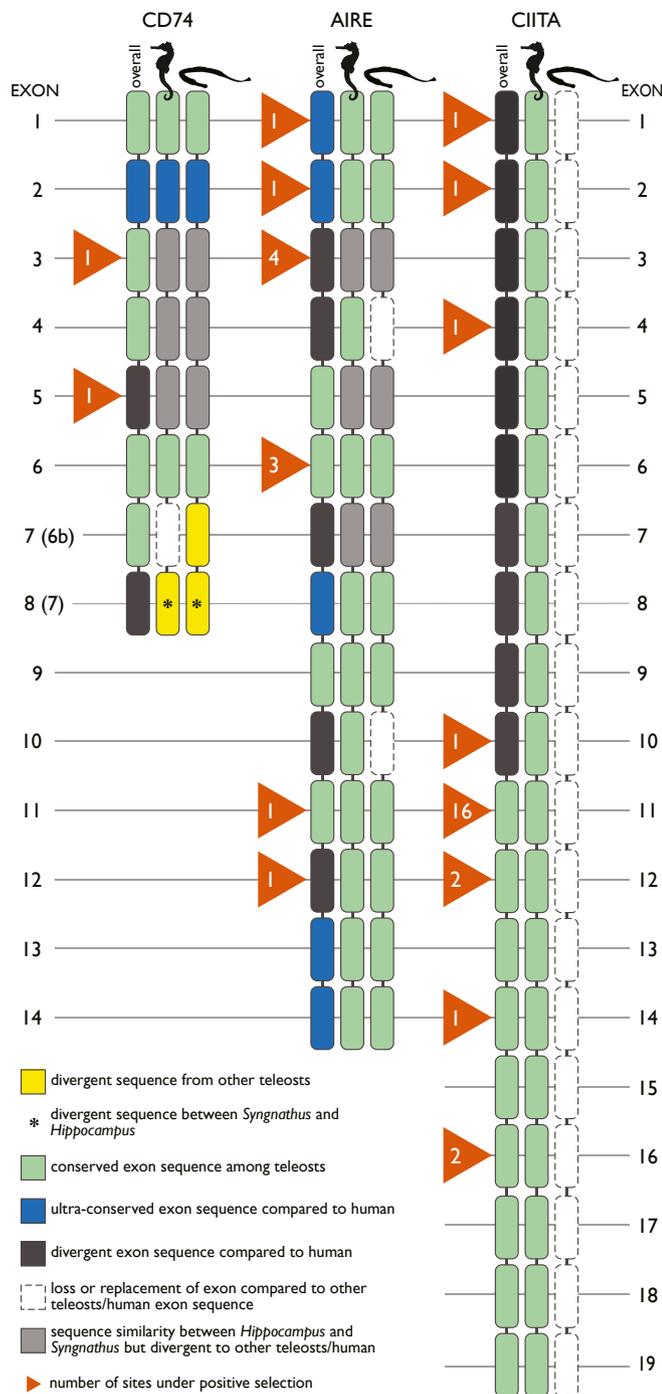
in the *Syngnathus* for exon 6b, accompanied with a divergent exon 3 in both lineages. While subsequently several core genes of the MHC II pathway were lost in *Syngnathus*, genes of the MHC II pathway were under positive selection in *Hippocampus* [Fig. 3 and SI Appendix, Table S11 and Dataset 1, gene alignments (31)] and showed clear sequence divergence compared to other teleosts and humans (Fig. 3). Different scenarios may explain the observed pattern in the MHC II pathway of *Hippocampus*.

First, the sequence divergence of the MHC II core genes in contrast to other teleosts and the signs of positive selection could indicate that in *Hippocampus* the MHC II genes were taking over alternative or novel functions. *CD74* is pivotal for a functional MHC II pathway, as supported by an impaired assembly and surface expression of *MHC II* and a defective antigen presentation in invariant chain knockout mice (37). While generally the CLIP of *CD74* (exon 3) associates with MHC II, the remaining exons of *CD74* act as chaperone, transporting MHC II to the loading compartment. The loss of exon 6b in *Hippocampus* could indicate a compromised loading process. Accordingly, the MHC II system in *Hippocampus* is likely to be less efficient in contrast to other vertebrates, which may suffice to permit the evolution of full male pregnancy.

Second, the MHC II pathway may not be compromised in its function despite the lost and diverged exons of *CD74* over a functional rearrangement of the immune system. However, this is less likely, as mice with transgenic expression of a truncated *CD74* protein lacking the CLIP region (the part of the gene that diverges from other teleosts in *Hippocampus*) could not pursue MHC II trafficking (38) (Fig. 3 and SI Appendix, Table S11).

### Modifications of the MHC I Pathway under Pregnancy

In Gadiformes (cod-like fishes) an independent loss of the MHC class II pathway was recently reported, and the observed diversification of *MHC I* genes was hypothesized to compensate for the loss of a functional MHC II pathway (39, 40). Accordingly, we assessed *MHC I* copy number in syngnathids using the most conserved exon 4 of the *MHC I* gene and found it to be higher in all species displaying male pregnancy (the *Nerophinae* with external male pregnancy [27 to 42 copies], *Hippocampus* [20 to 36 copies], and *Syngnathus* [24 to 44 copies] with full male pregnancy) compared to species without pregnancy (5 to 10 copies) (Fig. 2). While all identified *MHC I* sequences in Syngnathiformes are part of the U lineage (41), the distinct cluster of syngnathid *MHC I* sequences supports a potential coevolution of *MHC I* with male pregnancy (SI Appendix, Figs. S9 and S10). These lineage-specific *MHC I* variants likely increase the ligand repertoire and suggest a possible function within the cross-presentation pathway, in contrast to Atlantic cod, where cross-presentation could be hindered due to loss of the entire *CD74*, a gene with crucial function in the MHC I cross-presentation pathway (42). Moreover, key genes of the MHC I pathway, such as  $\beta$ 2-Microglobulin (*B2M*, important for the availability of *MHC I* light-chain proteins) and *CD8* (responsible for activation of  $CD8^+$  T lymphocytes), were under positive selection in syngnathids, similar to *RAG1* that facilitates V(D)J recombination and *TAP1/TAP2* that function as heterodimers in the transport of antigens (SI Appendix, Table S11). This supports a shift from the MHC II to MHC I cross-presentation pathway as all of the latter genes (*CD8*, *RAG1*, and *TAP1/TAP2*) also have important functions in the MHC I cross-presentation pathway. The



**Fig. 3.** Domain-level alignment of three genes (*CD74*, *AIRE*, *CIITA*) critical for the MHC class II pathway with lost or divergent exons in *Hippocampus* and *Syngnathus* compared to other vertebrate sequences. For each exon, sequence divergence to other teleosts (yellow), human (34), or sequence conservation other teleosts (green), human (blue) are shown. Exon loss is indicated in white and sequence conservation between *Hippocampus* and *Syngnathus* but divergence to other teleosts/humans in gray. Orange triangles indicate the number of sites under positive selection. An asterisk (\*) indicates yellow exon 6b and 7 of *CD74* also diverge between *Syngnathus* and *Hippocampus*.

identification of marked positive-selection signals support an interpretation of the MHC I pathway to coevolve with male pregnancy. The expanded MHC I repertoire is likely to be linked to the simultaneous loss/rearrangement of the MHC II pathway

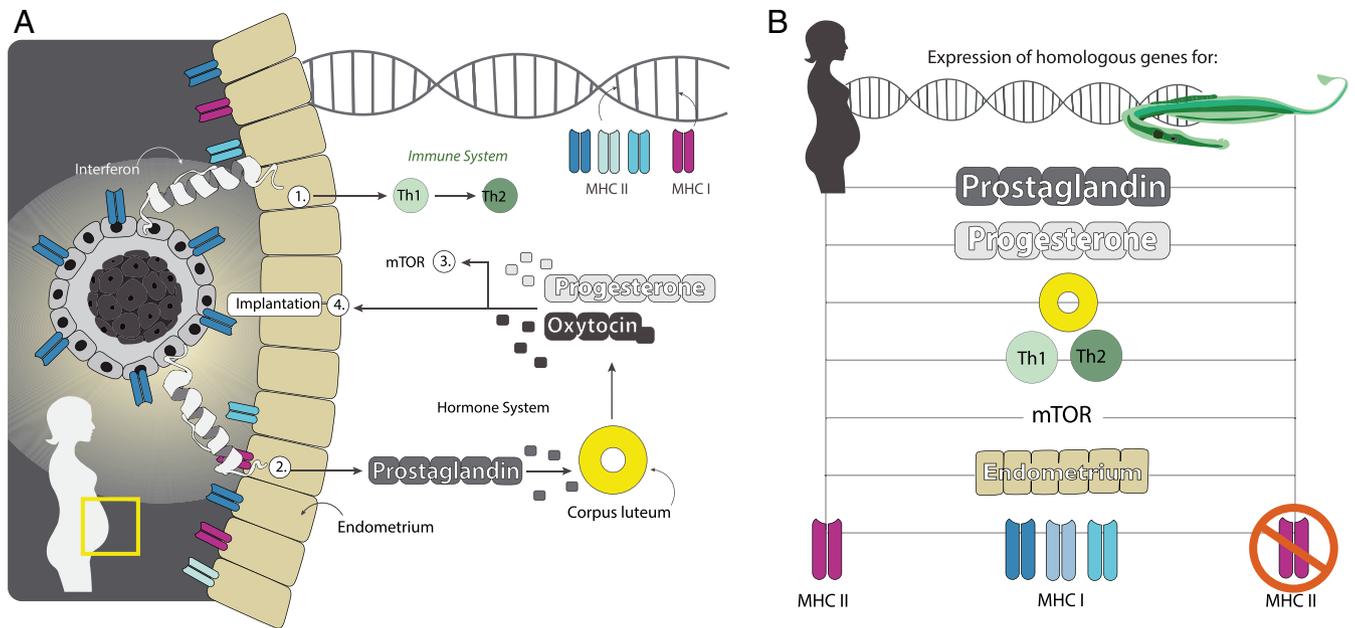
and may compensate its function/deficiency over the MHC I cross-presentation pathway.

Pregnancy requires special physiological adaptations to assure the oxygen supply to the growing embryo. In line with these expectations, the repertoire of hemoglobin genes encoding oxygen transport show signs of coevolution with male pregnancy. All syngnathids have lost the hemoglobin gene *alpha 6*, while those genera with full male pregnancy, *Syngnathus* and *Hippocampus*, have also lost the *alpha 5* gene. Conversely, fully pregnant species have gained *alpha 1* and *alpha 2* hemoglobin genes (SI Appendix, Figs. S14–S16). It is tempting to speculate that the shift in hemoglobin gene repertoire indicates selection for more effective oxygen transfer from father to offspring in male pregnancy evolution.

### Modulation of Gene Expression during Pregnancy

Next, we assessed whether or not the evolution of immunological tolerance required the cooption of similar genes and physiological processes in female and male pregnancy. To do so, we analyzed global gene-expression patterns using RNA sequencing in our model species *S. typhle* in brood pouch tissues during pouch development and pregnancy. At the same time, this approach assessed whether the evolution of immunological tolerance in mammals, in addition to the identified changes in gene repertoire. We examined the gene-expression profiles of male undeveloped brood pouch tissues (control) against developed pouch tissue from mature and receptive males (43); pouch tissue at early- and at late-pregnancy genes with a false-discovery rate-corrected  $P$  value of  $<0.05$ , as determined by the cuffdiff algorithm, were categorized as differentially expressed (44). All differentially expressed genes were searched for potential functions via homology, using reported functions in female pregnancy of mammals, in the squamate reptile *Chalcides ocellatus* (45) and in male pregnancy of *S. scovelli* (29) and *Hippocampus abdominalis* (46). A total of 141 genes were significantly up- or down-regulated during male pregnancy in *S. typhle* and *S. scovelli* (29). The direction of expression in differentially expressed genes correlated between *S. typhle* and *S. scovelli* ( $R^2 = 0.767$ ), implying that up- or down-regulation during pregnancy was mostly consistent in both pipefish species. In particular, this was the case for the four genes with the strongest up-regulation during pregnancy (*MYOC*, *HCEA*, *LS-12*, *APOA1*) and for the two genes that showed the most massive down-regulation during pregnancy (*STX2* and *MSXC*). Several genes known to be differentially expressed in the pregnancy of the seahorse *H. abdominalis* also showed expression changes in the pregnancy of the pipefish *S. typhle* (SI Appendix, Table S12). We identified 116 genes covering important pathways in human female pregnancy as differentially expressed during male pregnancy in *S. typhle*. These were involved in the prostaglandin, mammalian target of rapamycin (mTOR), and progesterone pathways, in corpus luteum degradation, parent-embryo transport, placenta development, conceptus implantation, and embryo growth (Fig. 4 and SI Appendix, section 7.3 and Tables S12 and S13).

In summary, these findings suggest that the independent evolutionary trajectories of female and male pregnancy in two vertebrate classes have resulted in expression changes in highly overlapping sets of genes coding for pathways with similar functions (SI Appendix, Tables S12 and S13). Apparently, the convergent evolution of male and female pregnancy has coopted a similar set of genes and involves similar physiological pathways. Functional tests using visualization of gene expression over in situ hybridization and gene knockdowns will permit further investigation of the molecular basis of male pregnancy evolution and its mechanistic similarity to female pregnancy in the future.



**Fig. 4.** Cooption of genes known from female human pregnancy in syngnathid male pregnancy. (A) Human female pregnancy and the most important immunological, developmental, and endocrinological pathways involved. (B) Genes involved in those pathways with an established role in human pregnancy change their expression also during pipefish pregnancy.

### Immune Gene Expression and Male Pregnancy

Next, we focused on immune gene-expression changes that accompany the modification of the MHC II pathway and the *MHC I* gene repertoire expansion. We analyzed the differential expression of immune genes that are either known to have a function in female pregnancy in mammals or known to also show expression changes in pregnancy of reptile, of seahorse, or of another pipefish species [*in bold italics* below; in the parenthesis *M* indicates a gene known to have a function in mammals, *R* in the reptile *C. ocellatus* (45), *S* in the seahorse *H. abdominalis* (46), and *P* in the pipefish *S. scovelli* (29)] (*SI Appendix, Table S12*) or those with a fold-change > 2 (220 genes in total, including 30 immune genes) (only in *italics*) (Fig. 5 and *SI Appendix, Table S13*).

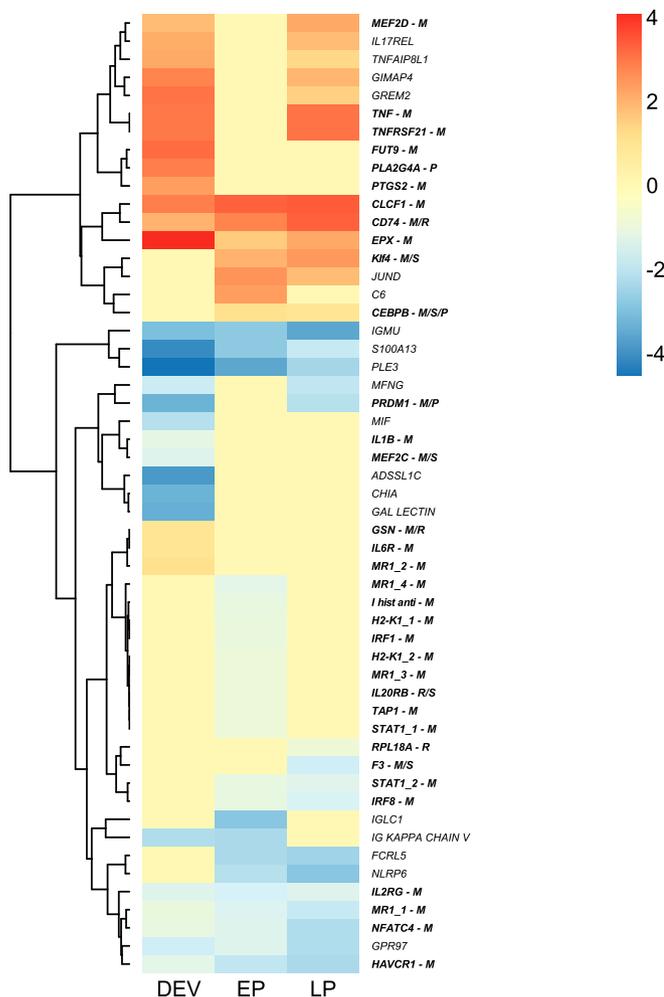
Collectively, the observed gene-expression changes during male pregnancy contribute to immunological tolerance during pregnancy already apparent from the gene repertoire. In particular, we identified expression changes of proinflammatory Th1 and antiinflammatory Th2 responses and a simultaneous down-regulation of the MHC I pathway during pregnancy, which resemble the expression changes during mammalian pregnancy. An inflammation response was suggested to be important for successful implantation in mammalian pregnancy (10). The key genes mediating this specific inflammation at implantation in mammals—*IL6R* (*M/P*), *TNF* (*M*), and *PTGS2* (*M*) (10, 47)—were up-regulated during pouch development in pipefish. Here, other inflammation responses dropped [down-regulation of proinflammatory interleukins *IL1B* (*M*) and *IL2RG* (*M*), and *S100A13* (interleukin secretion gene)]; the proinflammatory cytokine *MIF*; *FHL2* involved in inflammation response; *ADSSLIC* involved in antimicrobial peptide synthesis; the antimicrobial peptide *PLE3*; *JUND* involved in LPS response; and up-regulation of *GSN* (*M/R*) that binds to LPS].

Simultaneously, lymphocyte maturation and proliferation were suppressed through the down-regulation of *CH1A* and *MEF2C* (*M/S*) (maturation of B cells and important in mammalian embryo development), the up-regulation of *GIMAP4* that enhances lymphocyte apoptosis, and the up-regulation of the transcriptional

repressor *PRDM1* (*M/P*) that initiates in mammals a lineage-restricted progenitor cell population contributing to placental growth and morphogenesis (48). Consistent with a shift from Th1 to Th2 immune responses during mammalian pregnancy, *CEBPB* (*M/S/P*), which represses Th1 but facilitates Th2 immune response, was up-regulated during pipefish male pregnancy. This coincided with expression dynamics of *EPX* (*M*) mediating eosinophil activity and promoting mammalian placental development (49). Lymphocyte maturation and proliferation remained consistently repressed during pregnancy as indicated by down-regulations of *RPL18A* (*R*) (activation of T cell proliferation [Th1]), of *FCRL5* (enhancing B cell development), of the proinflammatory interleukin *IL2RG* (*R/S*), and of the interleukin secretion genes *S100A13* and *IL20R* (*R/S*).

During late pregnancy only, *GPR97* and *MFNG* (both responsible for B cell differentiation) were down-regulated along with the genes *NEATC4* and *HAVCR1*, which are involved in T cell maturation. Few genes involved in Th1 immune response were up-regulated during pregnancy [*TNF* (*M*), *CLCF1* (*M*), *KLF4* (*M/S*), and *TNFRSF21* (*M*)]. In female pregnancy, those genes were shown to have additional functions: *TNF* (*M*) mediates placental development and implantation (10, 50), *CLCF1* (*M*) is responsible for the onset of labors at term [a process resembling inflammation (51)], and *KLF4* (*M/S*) is key for the maintenance of gestation (52). The two inflammation genes, *PLA2G4A* (*P*) and *IL17REL*, were up-regulated during pouch development but not during pregnancy. In summary, inflammation responses during male pregnancy could be overlapping with previously identified expression patterns of homologous genes responsible for the regulation of inflammation during egg implantation and female pregnancy.

Analogous to human pregnancy where *CASP3* (*M/S*) modifies the MHC class I pathway (53, 54) to support immunological tolerance (55), *CASP3* (*M/S*) was up-regulated during pipefish pregnancy. Throughout early mammalian pregnancy, *TAP1* (*M*) is increasingly expressed on placenta-specific trophoblasts and plays an important role in preventing maternal immune attacks toward the embryos (56). Such up-regulation of *TAP1* (*M*) was



**Fig. 5.** Heatmap displaying expression changes ( $\log_2$  fold-change) of *S. typhle* immune genes during pouch development (DEV), early pregnancy (EP), and late pregnancy (LP). Homologs of genes marked with (M) possess known function (**bold**) in mammals (M), designation (R) denotes differential expression in the squamate reptile *C. ocellatus* (R), in the seahorse *H. abdominalis* (S), or in the pipefish species *S. scovelli* (P).

also identified in pipefish. Mammalian trophoblasts only express nonclassical nondiverse *MHC I* genes that will not induce a nonself-rejection reaction against the embryo (57, 58). In pipefish, a series of *MHC I* pathway genes that are involved in antigen recognition, presentation, and processing were down-regulated during pregnancy, such as *F10* (M), *H2-K1* (M), *IRF1* (M), *IRF8* (M), *MRI* (M), *FUT9* (M), and the Ig chains (*Ig  $\kappa$  chain V*, *Ig  $\mu$  chain C*, *IGLC1*). In humans, the silencing of *CD74* (M/R) during pregnancy is key for maintaining the acceptance of the semi-allogenic embryo (59). The up-regulation of *CD74* (M/R) is puzzling as almost all other genes of the *MHC II* pathway are absent in *S. typhle*. As key exons of *CD74* are diverged or substituted in *Syngnathus*, the up-regulation of *CD74* during pregnancy rather suggests a change of function for *CD74* in the evolution of male pregnancy. This suggests that consistent with female pregnancy, antigen recognition over the *MHC I* pathway could also be down-regulated during pipefish male pregnancy.

## Discussion

Although pregnancy is widespread among the vertebrates, very little is known on the immunological modifications that are required to prevent embryo rejection other than within the class

mammalia. Here, we present a major modification of the immune system associated with increasing investment into pregnancy in the fish family of pipefishes and seahorses that not only entailed gene-expression changes during pregnancy but also coincided with major alterations of the gene repertoire of both *MHC* pathways. While the identified rearrangement and loss of the core genes of the *MHC II* pathway is consistent with an adaptive explanation to modulate the immune system so as to prevent immunological rejection of the embryo, demonstrating causality would require future functional validation. While in *Syngnathus* the genomic knockout of the *MHC II* pathway must have resulted in a loss-of-function, in *Hippocampus* the situation remains inconclusive. Knockdowns in closely related teleost species of the *CD74* exon that was substituted in *Hippocampus* would be needed to illuminate its impact on the *MHC II* pathway.

One of the most unexpected findings was that even within a single fish family, the rearrangement of the *MHC II* pathway differed between the genera *Hippocampus* and *Syngnathus*. At the same time, this demonstrates both a strong selection for reduction of immunological vigilance displayed by the *MHC* class II pathway during pregnancy evolution and a remarkable flexibility of the vertebrate immune system in general. Because this unique fish family displays male pregnancy, any of the immunological adaptations are also not compounded by the sex per se (60) [i.e., the fact that the female sex through provisioning of eggs usually needs a more competent immune system under conventional sex roles, referred to as Bateman's principle (61)].

Within the evolution of vertebrate immune systems, the crucial role of the *MHC II* pathway for the recognition of pathogenic epitopes is commonly considered essential (8). The almost complete loss of the *MHC II* pathway in *Syngnathus* emphasizes that the vertebrate immune system has a much higher degree of functional flexibility than previously assumed, in line with recent findings in the Gadiformes lineage (34, 39). The complete loss of classic *MHC II*  $\alpha$  and  $\beta$  genes in all cod-like fishes (Gadiformes) coincides with truncation of *CD4* and loss of *CD74* (39, 40). While in that taxonomic group, the selection regime leading to *MHC* class II pathway loss is still elusive, we provide evidence among *Syngnathiformes* that modification and loss of adaptive immune genes and pathways is associated with the evolution of male pregnancy, which potentially selected for immunological tolerance. As *Syngnathids* and *Gadiformes* are only distantly related (*SI Appendix*, Fig. S1), losses and divergence of key genes of the *MHC II* pathway in each of those groups represent independent evolutionary events, likely driven by different selection factors.

The loss of gut-associated lymphatic tissues (GALT), the spleen (62), and the immune genes (*CD4*, *MHC II*, *AICDA*, *CIITA*) in *Syngnathus* represent critical pathways that are attacked by the HIV (CD4<sup>+</sup> T cells, GALT). As a natural “knockout” for the *MHC* class II pathway, *Syngnathus* may thus become instrumental in the future as a model for research on natural or disease-related immune deficiencies.

## Materials and Methods

We have sequenced and assembled 12 *Syngnathiformes* genomes (*SI Appendix*, Table S2) and annotated the genome of *S. typhle*. We generated a time-calibrated phylogeny of *Syngnathiformes* (*SI Appendix*, section 3 and Fig. S1). To search for shifts in the optima of genome size in the different lineages, we applied the Ornstein–Uhlenbeck process using the *Syngnathiformes* phylogeny and the genome sizes of the species (*SI Appendix*, section 4.1 and Fig. S2). To investigate potential reasons for differences in genome size, a library of repeated elements was created (*SI Appendix*, section 4.2 and Table S4). For immune, pregnancy, and hemoglobin genes, translated query sequences, either as whole sequences and for *MHC* and hemoglobin also split into individual exons, were used as input in a TBLASTN search toward the scaffold from the assembled draft genomes. For *MHC I* and *MHC II* searches, unigigs were used due to large copy numbers (*SI Appendix*, section

5.1). Upon gene alignments, gene trees were generated with RAxML (v8.2.10.) (SI Appendix, section 5.2) and local gene synteny was explored for *MHC II*, *AID*, *CD4*, and *CIITA* (SI Appendix, section 5.4). We assessed site-specific, positive selection and gene-wide selection across the whole tree and with the syngnathids as the foreground branches (SI Appendix, section 6 and Table S11). To assess which genes are differentially expressed during male pregnancy and during the development of the brood pouch tissue, we sequenced the transcriptome of *S. typhle* in different tissue types (undeveloped brood pouch, developed pouch, pouch at early pregnancy, pouch at late pregnancy). Differential gene expression was calculated pairwise against the undeveloped pouch and all differentially expressed genes were searched for potential functions via homology (SI Appendix, section 7).

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